

Genome Sequence of *Sphingomonas* sp. Strain PAMC 26621, an Arctic-Lichen-Associated Bacterium Isolated from a *Cetraria* sp.

Hyungseok Lee,^a Seung Chul Shin,^a Jungeun Lee,^a Su Jin Kim,^b Bum-Keun Kim,^c Soon Gyu Hong,^a Eun Hye Kim,^a and Hyun Park^a

Korea Polar Research Institute, Yeosu-gu, Incheon, South Korea^a; College of Life Sciences and Biotechnology, Korea University, Seongbuk-gu, Seoul, South Korea^b; and Korea Food Research Institute, Bundang-gu, Sungnam, South Korea^c

The lichen-associated bacterial strain *Sphingomonas* sp. PAMC 26621 was isolated from an Arctic lichen *Cetraria* sp. on Svalbard Islands. Here we report the draft genome sequence of this strain, which could provide novel insights into the molecular principles of lichen-microbe interactions.

Recently, a few studies have characterized the bacterial communities of lichens by culture-independent methods like next-generation sequence (NGS) analysis (4, 7, 8), suggesting the importance of the symbiotic relationship between lichen and lichen-associated microbes. *Sphingomonas* was identified as one of the dominant Gram-negative taxa inside several lichen species (4). Members of the genus *Sphingomonas* are traditionally known as degraders well adapted for the bioremediation of polycyclic aromatic hydrocarbons (3, 6, 10), and *Sphingomonas* has some oligotrophic strains isolated from extreme environments such as Antarctic soil (2, 10). In this study, the genome sequence of *Sphingomonas* sp. strain PAMC 26621, which was isolated from an Arctic lichen *Cetraria* sp. that grows on rocks (78°54'73"N, 11°57'78"E), was determined to provide novel insight into the molecular principles of lichen-microbe interactions.

The genome of *Sphingomonas* sp. PAMC 26621 was analyzed using a combined approach with the 454 GS FLX Titanium system (Roche Diagnostics, Branford, CT) with an 8-kb paired-end library (90,068 reads) and the Illumina GAIIX (San Diego, CA) with a 500-bp paired-end library (15,486,626 reads). The 454 GS FLX sequencing achieved about 3.5-fold coverage, while 290.2-fold read coverage was achieved by Illumina paired-end sequencing. The reads generated by the Illumina GAIIX and the 454 GS FLX Titanium were assembled using Celera Assembler 7.0 (9). Gene prediction and annotation were carried out using Glimmer3 (5), the RAST annotation server (1), and the NCBI COG database (11). The draft genome sequence of *Sphingomonas* sp. PAMC 26621 includes 4,769,913 bases and comprises 4,817 predicted coding sequences. It consists of 68 contigs (N_{50} contig size was approximately 210 kb), which can be assembled into eight scaffolds (N_{50} scaffold size was approximately 2,722 kb). The G+C content was 65.3%. Additionally, 47 tRNA-encoding genes, 2 23S rRNA genes, and 2 16S rRNA genes were predicted in the draft genome. Approximately 87.7% of nucleotides were predicted as protein-coding regions, and 67.4% (3,246) of the open reading frames were annotated with known proteins. Comparison with genome sequences available at RAST showed that *Sphingomonas* sp. strain SKA58 (score, 542), *Sphingopyxis alaskensis* RB2256 (score, 540), and *Sphingobium japonicum* UT26S (score, 496) were the closest neighbors of *Sphingomonas* sp. strain PAMC 26621.

By determining the genome sequence of *Sphingomonas* sp. PAMC 26621, it is now possible to perform various comparative genomic analyses with other bacteria, which should provide clues regarding the functional roles of this microbe within the lichen symbiosis.

Nucleotide sequence accession numbers. This Whole Genome Shotgun project has been deposited at DDBJ/EMBL/GenBank under the accession number [AIDW000000000](https://www.ncbi.nlm.nih.gov/nuccore/AIDW000000000). The version described in this paper is the first version, AIDW01000000.

ACKNOWLEDGMENT

This work was supported by a Functional Genomics on Polar Organisms grant (PE12020) funded by the Korea Polar Research Institute (KOPRI).

REFERENCES

1. Aziz RK, et al. 2008. The RAST Server: rapid annotations using subsystems technology. *BMC Genomics* 9:75.
2. Balkwill DL, et al. 1997. Taxonomic study of aromatic-degrading bacteria from deep-terrestrial-subsurface sediments and description of *Sphingomonas aromaticivorans* sp. nov., *Sphingomonas subterranea* sp. nov., and *Sphingomonas stygia* sp. nov. *Int. J. Syst. Bacteriol.* 47:191–201.
3. Baraniecki C, Aislabie J, Foght J. 2002. Characterization of *Sphingomonas* sp. Ant 17, an aromatic hydrocarbon-degrading bacterium isolated from Antarctic soil. *Microb. Ecol.* 43:44–54.
4. Bates ST, Cropsey GW, Caporaso JG, Knight R, Fierer N. 2011. Bacterial communities associated with the lichen symbiosis. *Appl. Environ. Microbiol.* 77:1309–1314.
5. Delcher AL, Harmon D, Kasif S, White O, Salzberg SL. 1999. Improved microbial gene identification with GLIMMER. *Nucleic Acids Res.* 27:4636–4641.
6. Harayama S. 1997. Polycyclic aromatic hydrocarbon bioremediation design. *Curr. Opin. Biotechnol.* 8:268–273.
7. Hodgkinson BP, Gottel NR, Schadt CW, Lutzoni F. 2012. Photoautotrophic symbiont and geography are major factors affecting highly structured and diverse bacterial communities in the lichen microbiome. *Environ. Microbiol.* 14:147–161.
8. Mushegian AA, Peterson CN, Baker CC, Pringle A. 2011. Bacterial diversity across individual lichens. *Appl. Environ. Microbiol.* 77:4249–4252.
9. Myers EW, et al. 2000. A whole-genome assembly of *Drosophila*. *Science* 287:2196.
10. Shi T, Fredrickson JK, Balkwill DL. 2001. Biodegradation of polycyclic aromatic hydrocarbons by *Sphingomonas* strains isolated from the terrestrial subsurface. *J. Ind. Microbiol. Biotechnol.* 26:283–289.
11. Tatusov RL, et al. 2001. The COG database: new developments in phylogenetic classification of proteins from complete genomes. *Nucleic Acids Res.* 29:22–28.

Received 13 March 2012 Accepted 22 March 2012

Address correspondence to Hyun Park, hpark@kopri.re.kr.

Copyright © 2012, American Society for Microbiology. All Rights Reserved.

doi:10.1128/JB.00395-12