

Functional Analysis and Enzymatic Modification by Site-Directed Mutagenesis of an Omega-6 Fatty Acid Desaturase from Arctic *Chlamydomonas* sp.

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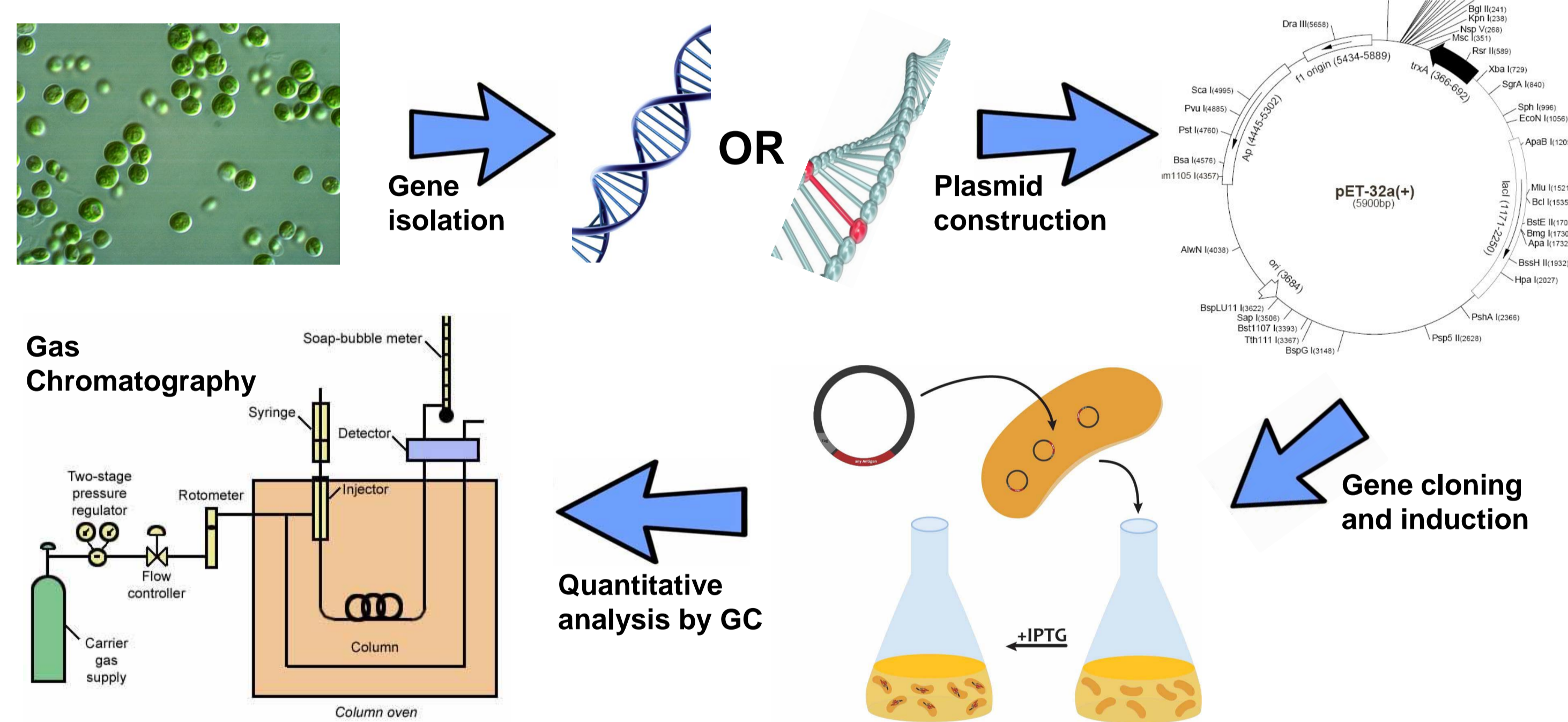
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ABSTRACT

Arctic *Chlamydomonas* sp. is a dominant microalgal strain in cold or frozen freshwater in the Arctic region. The full-length open reading frame of the omega-6 fatty acid desaturase gene (AChFAD) was obtained from the transcriptomic database of Arctic *Chlamydomonas* sp. from the KOPRI Culture Collection of Polar Microorganisms (KCCPM). Amino acid sequence analysis indicated the presence of three conserved histidine-rich segments as unique characteristics of omega-6 FADs, and three transmembrane regions transported to plastidic membranes by chloroplast transit peptides in the N-terminal region. Arctic *Chlamydomonas* sp. omega-6 fatty acid desaturase with 48.2 kDa showed enzymatic activity enhancing the concentration of linoleic fatty acid in the *E. coli* expression system. The AChFAD6 desaturase activity was examined by expressing wild-type and V254A mutant (Mut-AChFAD6) heterologous recombinant proteins. Quantitative gas chromatography indicated that the concentration of linoleic acids in AChFAD6-transformed cells increased more than three-fold (6.73 ± 0.13 mg g⁻¹ dry cell weight (DCW)) compared with cells transformed with vector alone. In contrast, transformation with Mut-AChFAD6 increased the concentration of oleic acid to 9.23 ± 0.18 mg g⁻¹ DCW, indicating a change in enzymatic activity to mimic that of stearoyl-CoA desaturase (SCD). These results demonstrate that AChFAD6 of Arctic *Chlamydomonas* sp. increases membrane fluidity by enhancing desaturating C18 fatty acids and facilitates production of large quantities of linoleic fatty acids in prokaryotic expression systems. Therefore, genetic modification on the membrane-spanning regions of fatty acid desaturases might increase the yield of target fatty acids and modulate enzymatic activities for industrial application.

MATERIALS & METHODS



RESULTS

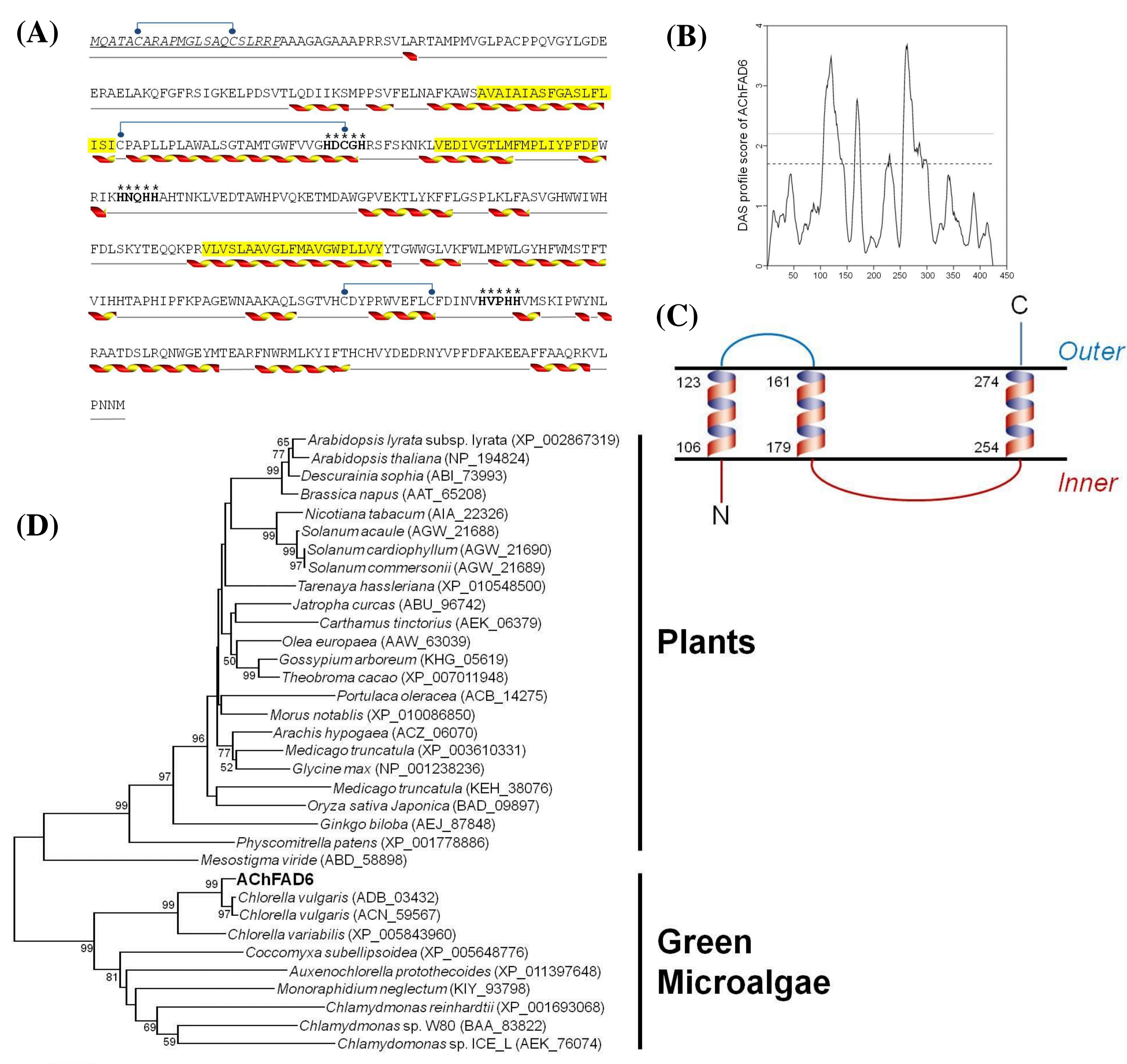


Fig. 1. Information of the secondary structure (A), membraneous topology (B and C) and phylogenetic relationship (D) of Arctic *Chlamydomonas* sp. omega-6 fatty acid desaturase (FAD, AChFAD6). In the figure (A), blue lines, yellow highlights and asterisks indicate disulfide bridges, transmembrane amino acids and Histidine-rich motifs, respectively.

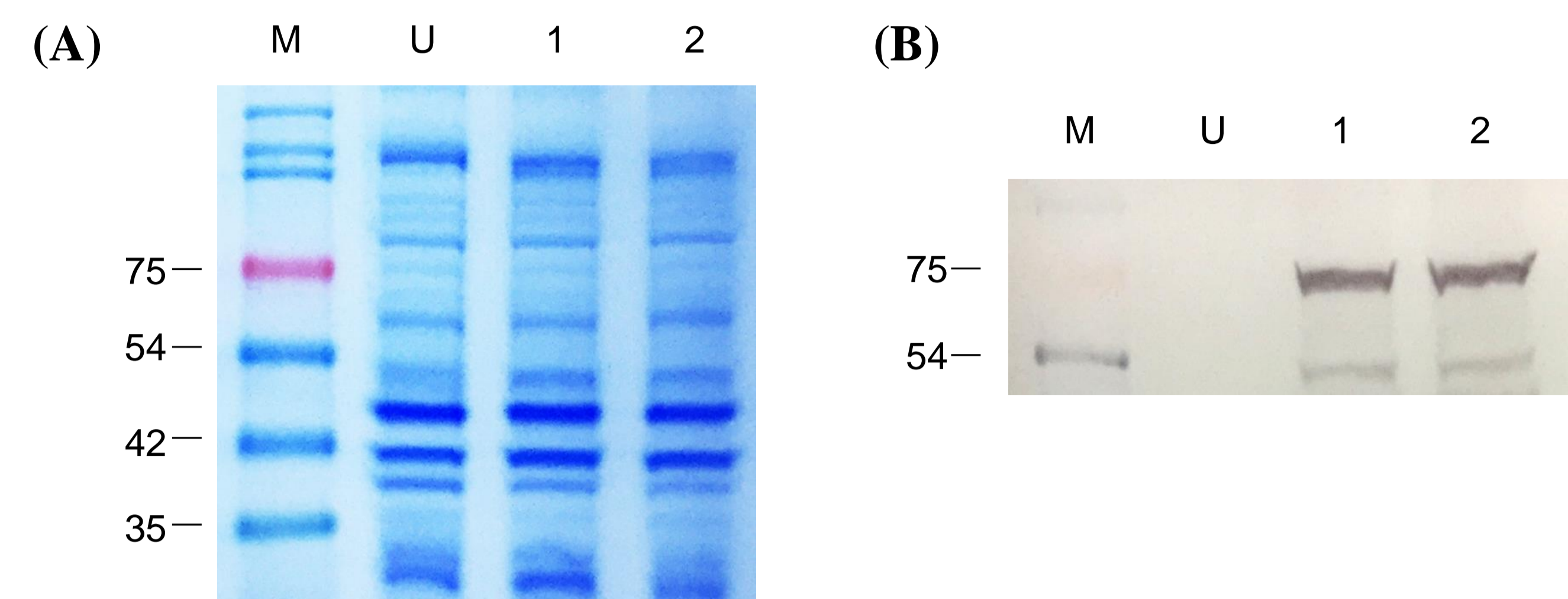


Fig. 2. Heterologous expression and detection of recombinant AChFAD6 in *E. coli*. M, protein ladder; U, Uninduced cells (Vector only); 1 and 2, *E. coli* cells harboring AChFAD6 and Mut-AChFAD6, respectively

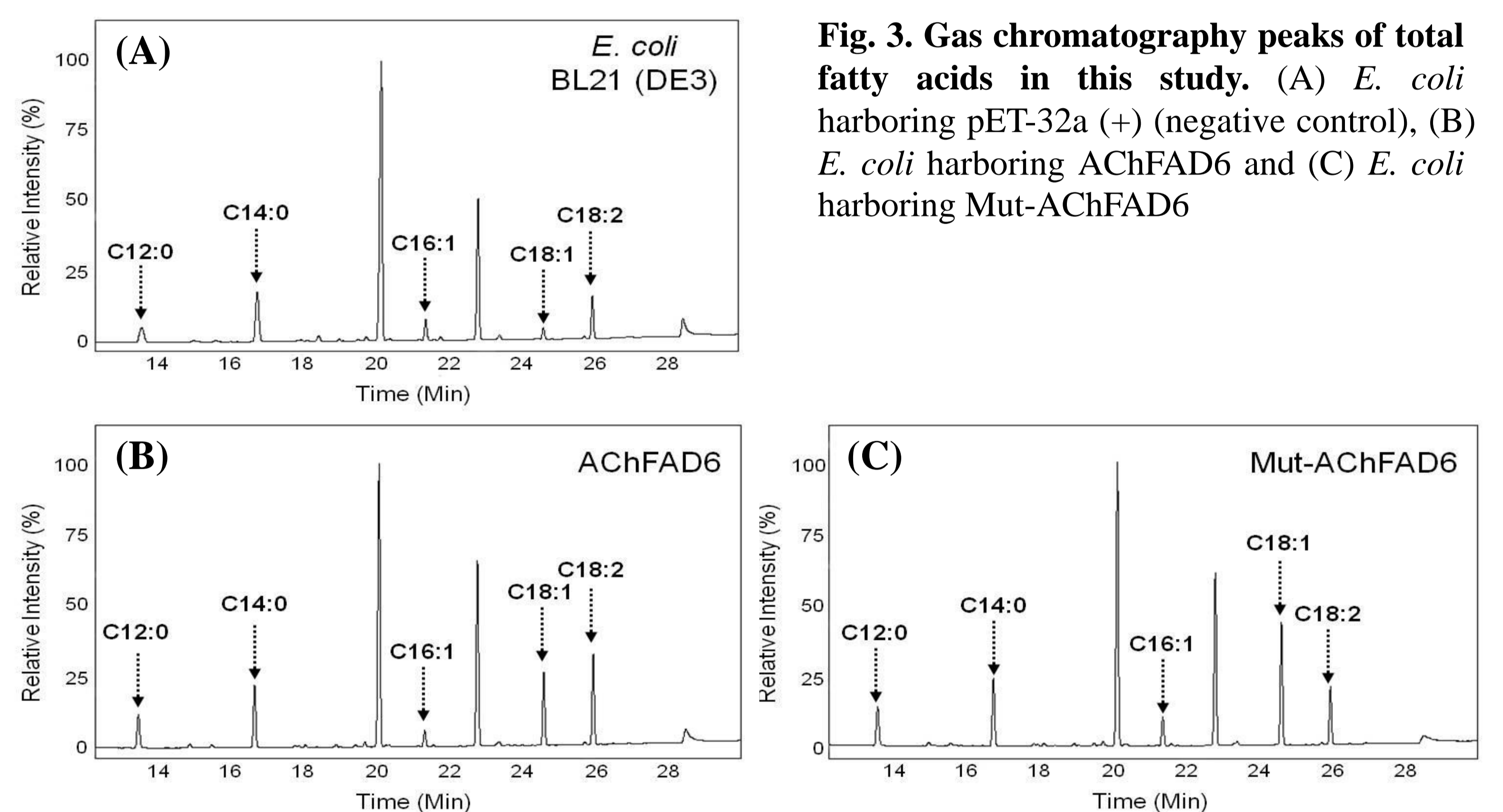


Fig. 3. Gas chromatography peaks of total fatty acids in this study. (A) *E. coli* harboring pET-32a (+) (negative control), (B) *E. coli* harboring AChFAD6 and (C) *E. coli* harboring Mut-AChFAD6

Table 1. Dominant contents of fatty acid methyl esters (FAMES) in expressed *E. coli* cells modified by induction of AChFAD6 and Mut-AChFAD6

	Organisms analyzed			
	<i>E. coli</i> (BL21(DE3))	AChFAD6	Mut-AChFAD6	<i>C. reinhardtii</i> cc-125
C12:0 (mg/g, %)	1.54 ± 0.05 (4.06 ± 0.24%)	2.99 ± 0.17 (5.32 ± 0.97%)	3.48 ± 0.17 (5.39 ± 0.35%)	-
C14:0 (mg/g, %)	3.85 ± 0.15 (10.12 ± 0.65%)	5.02 ± 0.25 (8.93 ± 1.58%)	5.38 ± 0.21 (8.32 ± 0.45%)	-
C16:1 (mg/g, %)	1.11 ± 0.06 (2.92 ± 0.23%)	1.26 ± 0.05 (2.24 ± 0.38%)	2.23 ± 0.07 (3.45 ± 0.16%)	1.00 ± 0.02 (0.77 ± 0.03%)
C18:1 (mg/g, %)	0.56 ± 0.05 (1.47 ± 0.17%)	5.45 ± 0.16 (9.67 ± 1.52%)	9.23 ± 0.18 (14.27 ± 0.49%)	1.40 ± 0.04 (1.08 ± 0.05%)
C18:2 (mg/g, %)	2.06 ± 0.12 (5.41 ± 0.45%)	6.73 ± 0.13 (11.92 ± 1.76%)	4.24 ± 0.13 (6.56 ± 0.30%)	-
Total FAs (mg/g)	38.07 ± 0.97	63.66 ± 1.27	64.71 ± 0.97	130.00 ± 1.56

* FAME composition: C12:0, Lauric acid methyl ester; C14:0, Myristic acid methyl ester; C16:1, Palmoleic acid methyl ester; C18:1, Oleic acid methyl ester; C18:2, Linoleic acid methyl ester; -: not detected

CONCLUSION

- The gene and the deduced amino acid sequences of omega-6 fatty acid desaturase (FAD) were isolated and enzymatic activity was investigated by gas chromatography.
- The omega-6 FAD with 48.2 kDa showed typical activity of the enzyme improving the concentration of linoleic fatty acid in the *E. coli* expression system.
- An V254A as substitution of the first amino acid in the third transmembrane domain resulted in an enzymatic change from FAD to stearoyl-CoA desaturase.
- The first amino acid in third membrane-spanning region is crucial to maintain omega-6 FAD activity. Therefore, genetic modification on the membrane-spanning regions of FADs can increase the yield of target fatty acids and modulate enzymatic activities for industrial application.

ACKNOWLEDGEMENT

This project was supported by the basic research program (PE16020) of the Korea Polar Research Institute (KOPRI) and the KOPRI Project (PM15040) of Ministry of Oceans and Fisheries (MOF)