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# MSK2018

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#### Poste

#### B001

#### Artificial Warming Effect on Microbial Community and Humic Substance Degradation in Maritime Antarctic Soil in King George Island

Ha Ju Park<sup>1</sup>, Jung Ho Kim<sup>2</sup>, Ui Joung Youn<sup>1</sup>, Angélica Casanova-Ka<sup>3</sup>, Soon Gyu Hong<sup>1</sup>, and <mark>Dockyu Kim<sup>1</sup>\*</mark>

<sup>1</sup>Division of Polar Life Sciences, Korea Polar Research Institute, <sup>2</sup>Department of Microbial Engineering, Konkuk University, <sup>3</sup>School of Environmental Science, Catholic University of Temuco

#### B002

Endophytic Bacteria from Turmeric Plant as Plant Growth Promoter and Antifungal Agents

Min-Ok Jun<sup>1,2</sup>, Cheol-Hee Kim<sup>2</sup>, Suk Weon Kim<sup>1</sup>\*, and Jiyoung Lee<sup>1</sup>\* <sup>1</sup>Korea Research Institute of Bioscience & Biotechnology (KRIBB), <sup>2</sup>Department of Biology, Chungnam National University

#### B004

Comparison of Microbiome and Virome in Chicken Farm under Diverse Environmental Factors

Jongbin Park<sup>1</sup>, Gwi-Deuk Jin<sup>1</sup>, Inhwan You<sup>1</sup>, Soo Jin Kim<sup>1</sup>, Sukjung Choi<sup>1</sup>, and Eun Bae Kim<sup>1,2,3</sup>\*

<sup>1</sup>Department of Animal Life Science, Kangwon National University, <sup>2</sup>Division of Applied Animal Science, Kangwon National University, <sup>3</sup>Institute of Animal Resources, Kangwon National University

#### B005

Microbial Communities Responsible for Glucose and Acetate in Organic-enriched Sediment of the Fin-fish Farm in the South Coast of Korea

Ayeon Choi<sup>1</sup>, Hyeyoun Cho<sup>1</sup>, Hyun-Ji Lee<sup>1</sup>, Hyun-Soo Beak<sup>1</sup>, Ju-Wook Baek<sup>1</sup>, Ui-jung Jung<sup>1</sup>, SungUk An<sup>1</sup>, Hyung Chul Kim<sup>2</sup>, Won-Chan Lee<sup>2</sup>, and Jung-Ho Hyun<sup>1</sup>\*

<sup>1</sup>Department of Marine Science and Convergent Technology, Hanyang University, <sup>2</sup>Marine Environment Research Division, National Institute of Fisheries Science (NIFS)

#### B006

#### Metagenomic Study of the Upper Airway Microbiome Associated with Childhood Asthma

Min-Jung Lee, Doo-heon Son, and Bong-Soo Kim\* Department of Life Sciences, Multidisciplinary Genome Institute, Hallym University

#### B007

Synergistic Effect of Antimicrobial Substances from Some Bacteria against Human Skin Pathogens

Da-Sol Lee and Hong-Gyu Song\* Department of Biological Sciences, Kangwon National University

#### B008

Aflatoxin B<sub>1</sub> Biodegradation and Inhibition against Aflatoxigenic Fungi by *Streptomyces* spp.

Ji-Seon Hwang and Hong-Gyu Song\* Department of Biological Sciences, Kangwon National University

#### B009

Bacterial Degradation of Ochratoxin A and Growth Inhibition of Ochratoxigenic *Aspergillus* spp.

Ho-Yeong Choi and Hong-Gyu Song\* Department of Biological Sciences, Kangwon National University

#### B010

#### Insight the Genome of *Lacinutrix venerupis* Strain DOK2-8 Isolated from Dokdo, Republic of Korea

Se Ra Lim<sup>1,2</sup> and Ji Hyung Kim<sup>1</sup>\*

<sup>1</sup>Infectious Disease Research Center, Korea Research Institute of Bioscience and Biotechnology, <sup>2</sup>Bio-Analytical Science Division, University of Science and Technology

#### B011

#### An *in vitro* Study to Assess the Impact of Tetracycline on the Human Intestinal Microbiome

Ji Young Jung

Applied Bioresources Research Division, Freshwater Bioresources Utilization Bureau, Nakdonggang National Institute of Biological Resources (NNIBR)

#### B012

Characterization of Root-associated Microbiome Inhabiting in Different Compartments of Tomato Plants

Shin Ae Lee<sup>1</sup>, Yiseul Kim1, Jae-Ho Joa<sup>2</sup>, Bora Chu<sup>1</sup>, Mee Kyung Sang<sup>1</sup>, Jaekyeong Song<sup>1</sup>, and Hang-Yeon Weon<sup>1</sup>\*

<sup>1</sup>National Institute of Agricultural Sciences, <sup>2</sup>Research Institute of Climate Change and Agriculture, National Institute of Horticultural & Herbal Science



# Artificial Warming Effect on Microbial Community and Humic Substance Degradation in Maritime Antarctic Soil in King George Island

Ha Ju Park<sup>a</sup>, Jung Ho Kim<sup>b</sup>, Ui Joung Youn<sup>a</sup>, Angélica Casanova-Katny<sup>c</sup>, Soon Gyu Hong<sup>a</sup>, and Dockyu Kim<sup>a,\*</sup>

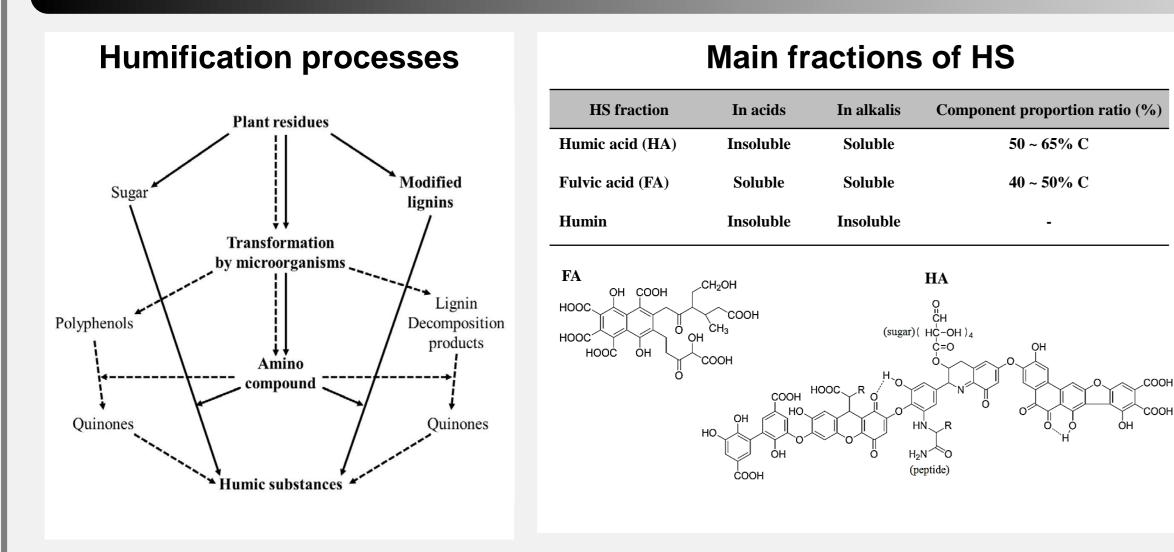
<sup>a</sup> Division of Polar Life Sciences, Korea Polar Research Institute, Incheon 21990, South Korea <sup>b</sup> Department of Microbial Engineering, Konkuk University, Seoul 143-701, South Korea <sup>c</sup> School of Environmental Science, Catholic University of Temuco, Rudecindo Ortega 02950, Temuco, Chile

## ABSTRACT

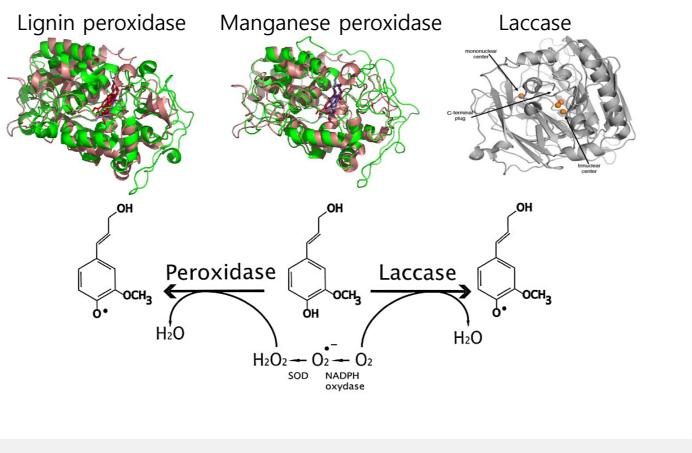
Although the maritime Antarctic has undergone rapid warming, the effects on indigenous soil-inhabiting microorganisms are not well known. Artificial warming experiments using open-top chamber (OTC) have been performed on the Fildes Peninsula in the maritime Antarctic since 2008. When the soil temperature was measured at a depth of 2–5 cm during the 2013–2015 summer seasons, the mean temperature inside OTC (OTC-In) increased by approximately 0.8°C compared with outside OTC (OTC-Out), while soil chemical and physical characteristics were not changed. Soils from OTC-In and OTC-Out were subjected to analysis for change in microbial community and degradation rate of humic substances (HS, the largest pool of recalcitrant organic carbon). Archaeal and bacterial communities in OTC-In were minimally affected by warming compared with those in OTC-Out, with archaeal methanogenic *Thermoplasmata* slightly increased in abundance. The abundance of heterotrophic fungi *Ascomycota* was significantly altered in OTC-In. Total bacterial and fungal biomass in OTC-In increased by 20% compared to OTC-Out, indicating that this may be due to increased microbial degradation activity for soil organic matter (SOM) including HS, which would result in the release of more lowmolecular-weight growth substrates from SOM. Despite the effects of warming on the microbial community over the 8-years-experiments warming did not induce any detectable change in content or structure of polymeric HS. This work was supported by grant to KOPRI (PE18090)

## INTRODUCTION

RESULTS

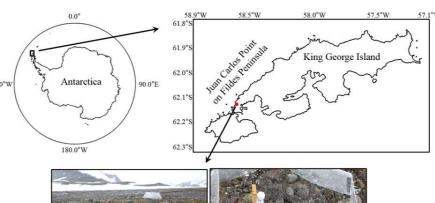


Nonspecific oxidizing enzymes for initial HS degradation (proposed)

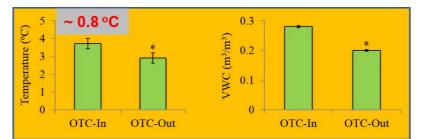


## METHODS

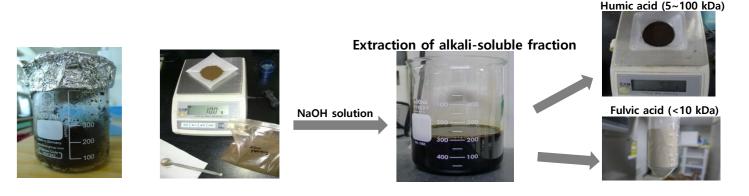
- Sampling site and open top chamber (OTC) installation











- NMR-based structural analysis of HA and FA
- Microbial community analysis (16S rRNA gene pyrosequencing & PLFA )

Table 1. Soil chemical and physical characteristics inside the OTCs (OTC-In) and in control plots (OTC-Out)

Sample	Chemical and physical characteristics of soil								
	EC (µs/m)	рН	% N	% C	C/N	% sand	% lime	% clay	
OTC-Out	59.6	6.5	0.094	0.968	10.3	71.7	23.7	4.5	
	(±16.8)	(±0.2)	(±0.021)	(±0.218)	(±0.4)	(±2.6)	(±2.1)	(±0.5)	
OTC-In	64.4	6.6	0.092	0.994	10.8	69.6	25.2	5.2	
	(±23.7)	(±0.3)	(±0.018)	(±0.171)	(±0.7)	(±3.1)	(±2.6)	(±0.5)	

The statistical data are the mean values of three independent analyses for each OTC-Out and OTC-In soil.

Table 2. Summary of pyrosequencing results and statistical analyses of microbial communities in soil samples from open-top chamber (OTC) passive warming experimental sites

Target microorganism		Number of reads				
			Observed	Chao1	ACE	OTU diversity (Shannon)
Bacteria	OTC-Out	8064±1030	1783±276	3016±422	4007±494	6.49±0.08
	OTC-In	6888±364	1917±399	3301±1022	4425±1632	6.74±0.24
Archaea	OTC-Out	5919±3883	34±10	39±11	44±14	1.48±0.48
	OTC-In	6167±4824	28±13	31±13	31±13	1.42±0.68
Fungi	OTC-Out	8826±2049	463±74	576±69	581±69	4.39±0.70
	OTC-In	9987±1006	401±25	524±49	505±40	3.98±0.05

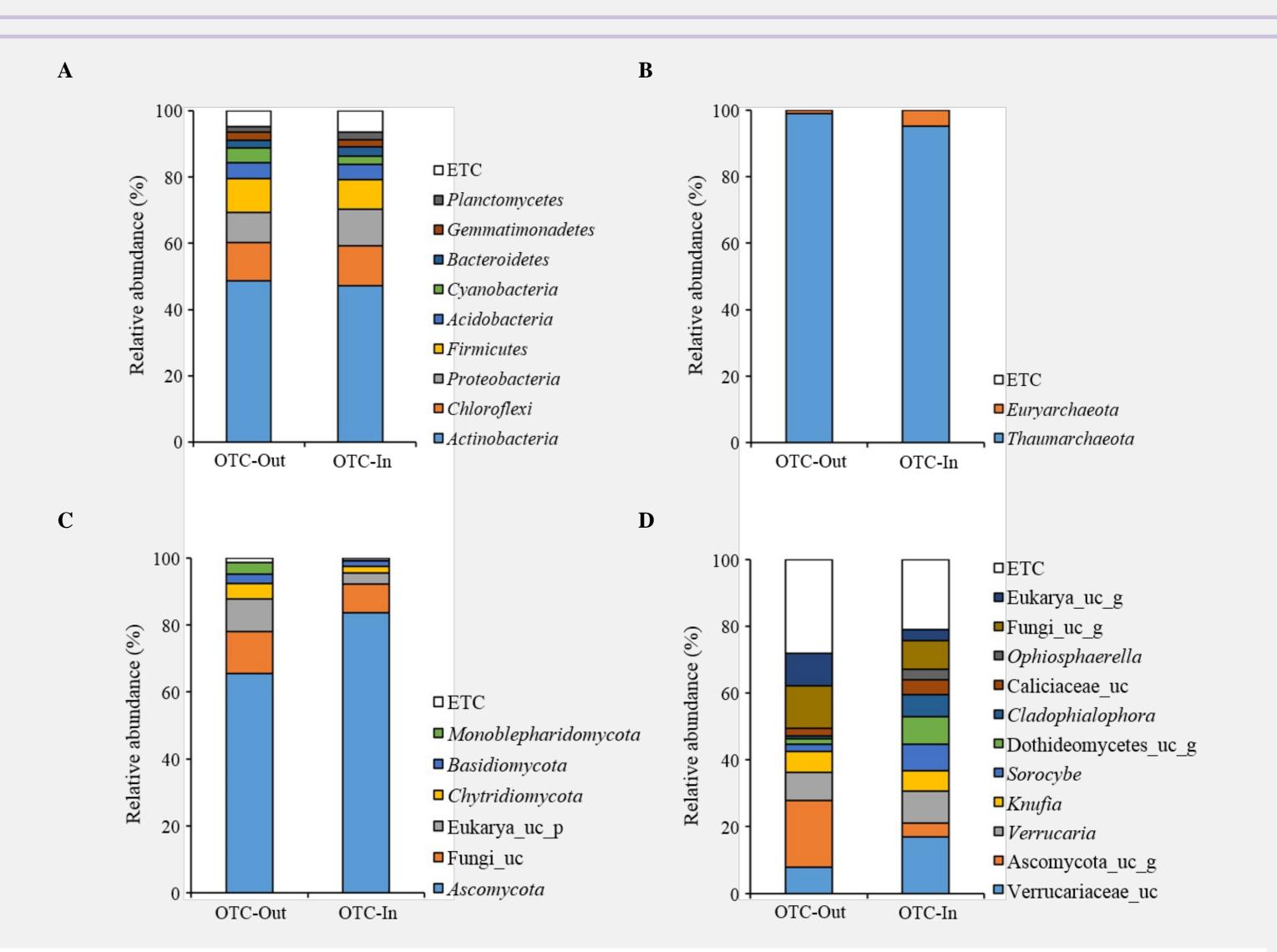


Fig. 1. Changes in abundance and taxonomic composition of soil microbes induced by the OTC warming experiment. Relative abundances of (a) bacterial phyla, (b) archaeal phyla, (c) fungal phyla, and (d) different genera within the fungal phylum *Ascomycota* are shown. ETC denotes the sum of minor taxa (< 1.0% in relative abundance). Relative abundances are the mean values of three independent analyses for each OTC-Out and OTC-In soil.

The statistical data are the mean values of three independent analyses for each OTC-Out and OTC-In soil.

Table 3. Quantity of fungal and bacterial phospholipid-derived fatty acids (PLFAs) in soil samples from open-top chamber (OTC) passive warming experimental sites

	Amount of phospholipid fatty acids (mg/kg soil)		
	OTC-Out	OTC-In	
Fungi	6.8	8.0	
Bacteria	5.8	7.5	
Gram+	3.2	4.4	
Gram-	1.8	2.0	
Actinomycete	0.8	1.1	
Total	12.6	15.5	
Ratio of fungal/bacterial biomass	1.17	1.07	

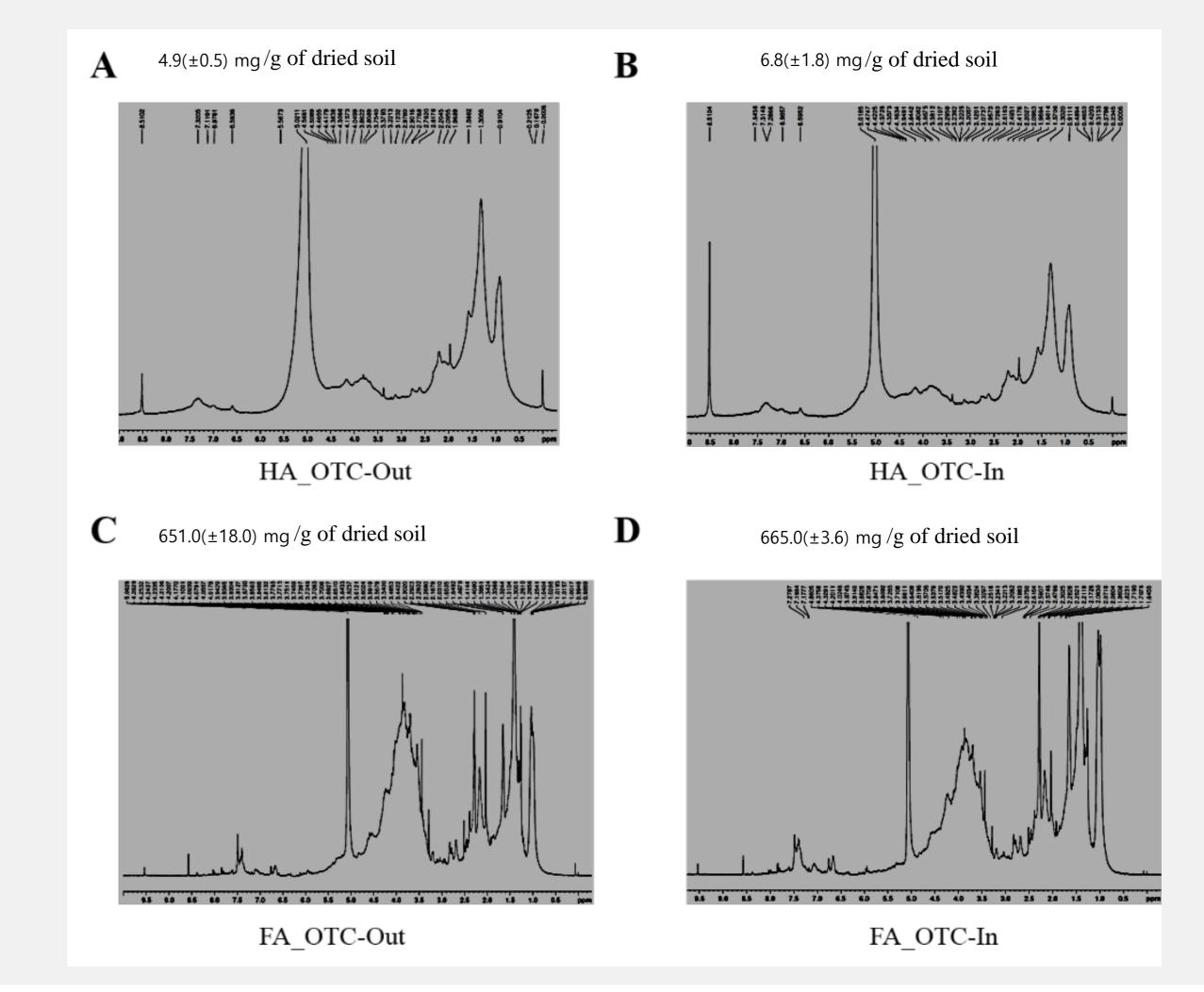


Fig. 2. <sup>1</sup>H-NMR spectral analysis of humic acid (HA) and fulvic acid (FA) in OTC warming experimental soils. (a) HA in OTC-Out; (b) HA in OTC-In; (c) FA in OTC-Out; (d) FA in OTC-In.