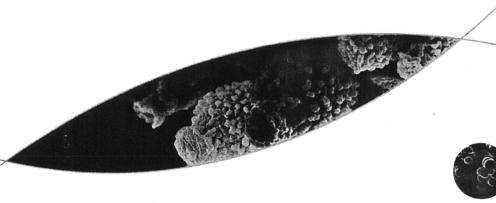
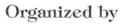
**Proceedings** 

# 2009 **International Meeting of** the Federation of Korean Microbiological Societies





Hotel-Seoul Kyoyuk Munhwa Hoekwan, Seoul, Korea



The Federation of Korean Microbiological Societies (FKMS) National Institute of Biological Resources

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### Characterization of Myxobacteria Extracts Against Biological Activity

Yongsub Yi1,2 and Jaeyoung Kim2

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Myxobacteria extracts were determined for application in cosmetic field. firstly, tyrosinase activity was determined and shown in No. 1952 having 20% inhibition. For screening of anti-bacteria activity, growth inhibition of *Staphylococcus aureus*, *Staphylococcus epidermidis*, and *E. coli* K12 were determined. The extracts didn't inhibited growth of *Staphylococcus aureus* and *Staphylococcus epidermidis*, but did the growth.

Keywords: biological activity, Myxobacteria, tyrosinase

### H054

# Biological Characterization of Glucosidase Isolated from Soil Bacteria

Yongsub Yi<sup>1,2</sup>\* and Jaeyoung Kim²

<sup>1</sup>Department of Herbal Cosmetic Science, Hoseo University, <sup>2</sup>Department of Biochemistry, Hoseo University

Glucosidase from previously isolated bacteria was determined and isolated. The glucosidase showed activity at congo red and active staining in native gel. Reactive product was determined in HPLC analysis, and compared with Rg3, reactive product control. reactive product showed Rg3 peak at the same time. From these results, glucosidase have biotransformation of Rb1 to Rg3, and shows typically biological activity of glucosidase.

Keywords: glucosidase, biotransformation, active staining

### H055

# Purification and Characterization of Cold-Active $\beta$ -N-Acetylglucosaminidase from Arctic Bacterium Pseudoalteromonase issachenkonii KOPRI 22<sup>-14</sup>

Dockyu Kim, Ha Ju Park, Il-Chan Kim, Soon Gyu Hong, Sung Hong Kum Lee, and Joung Han Yim\*

Polar BioCenter, Korea Polar Research Institute, Korea Ocean Research & Development Institute

136 marine bacteria showing chitinolytic activity were isolated from 47 sediment samples of Kara Sea, Arctic. Among them, a psychrotolerant strain (KOPRI 22718) was selected for its coldactivity at 5°C. The sequence analysis of the 16S rRNA affiliates KOPRI 22718 to Pseudoalteromonas isschenkonii. An exo-acting chitinase (~100 kDa) was homogeneously purified from the culture supernatant of KOPRI 22718 through ion exchange and gel filtration chromatography. The purified chitinase (W-Chi22718) exhibited the highest activity toward pNP-GlcNAc and produced GlcNAc monomer as end-product from chitin oligosaccharides (GlcNAc<sub>2</sub>-GlcNAc<sub>6</sub>), due to its  $\beta$ -N-acetylglucosaminidase activity. W-Chi22718 displayed chitinase activities at 0-37°C (optimal temperature of 30°C), and maintained its activities at pH 6.0-9.0 (optimal pH of 7.6). Interestingly, W-Chi22718 exhibited a relative activity of 13 and 35% at 0 and 10°C, respectively, in comparison to 100% at optimal 30℃, which is comparable with those of the previously characterized, cold-adapted, bacterial chitinases. Among the main cations and protein denaturing reagents, W-Chi22718 activity could be enhanced by K<sup>+</sup>, Ca<sup>2+</sup>, and Fe<sup>2+</sup>, but completely inhibited Cu<sup>2+</sup>and SDS.

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Keywords : Arctic; psychrotolerant, cold-active, exochitinase,  $\beta$ -N-acetylglucosaminidase

#### H056

### Noble Substrate Specificity of UDP-Galactose-4-Epimerase from *Escherichia coli*

Hye-Jung Kim and Pil Kim\*

Department of Biotechnology, Catholic University

Sugar isomerases have broad substrate specificity, i.e., ribose-5phosphate isomerase catalyzes not only phosphate sugar but also common sugar as substrate for isomerization. Natural sugar-4epimerase, in contrast, has been reported catalyzing only a nucleic acid activated substrate, which means its catalytic site has a space for UDP recognition. To understand substrate specificity of sugar-4-epimerase forward common sugars, UDP-galactose-4-epimerase (galE) from Escherichia coli, which catalyzes interconversion from UDP-galactose to UDP-glucose in the galactose metabolism in vivo, has been chosen. Since the GalE has been revealed x-ray crystal structure including catalytic residues and co-factor (NAD+) recognition site, it is considered suitable to protein engineering and evolution purpose for changed substrate recognition. The galE gene was cloned into a pET20b and expressed protein was purified using Ni-affinity chromatography. The purified GalE was found having a catalytic activity for D-galactose and the optimal temperature and pH were 35 °C and 7.5, respectively. Under the optimal condition, the  $K_m$  and  $k_{cat}$  values were exhibited 26.4 mM and 0.0155 min<sup>-1</sup> for D-galactose. When it was compared with kinetic parameter of UDP-galactose,  $K_m$  was lower 90 fold and  $k_{cat}$  was higher 3x106 fold. Among all D-aldohexose and D-ketohexose, galE indicated 4-epimerization activity for glucose, galactose, fructose, tagatose, psicose, and sorbose as substrates. The specific activity for galactose, glucose, psicose, sorbose, fructose and tagatose were 0.47, 0.32, 15.44, 2.08, 2.82 and 9.67 U/mg, respectively. Substrate specificity evolution of GalE based on 3D-structure analysis and protein engineering are under investigation for 4-epimerization of common sugars.

Keywords: 4-epimerization, common sugar, nucleic acid activated sugar